

TEXAS CHILDREN'S HOSPITAL

EVIDENCE-BASED OUTCOMES CENTER Fever Without Localizing Signs (0-60 Days Old)

Evidence-Based Guideline

<u>Definition</u>: An acute febrile illness (temperature ≥100.4°F [38°C]) with uncertain etiology after completion of a thorough history and physical examination. (1-3)

Etiology: The most common cause of fever without localizing signs (FWLS) is a viral infection. The challenge lies in the difficulty of distinguishing serious bacterial illness (SBI) from viral illness in neonates and early infancy. (4,5)

Inclusion Criteria:

- Age 0-60 days (Infants ≥35 weeks gestation)
- · Neonates and infants without underlying conditions
- Actual rectal temp ≥100.4°F (38°C) OR reported temp (axillary or rectal) of ≥100.4°F (38°C) in any time during the past 24 hours
- Neonates aged ≤21 days with bronchiolitis
- Neonates and infants (0-60 days) with respiratory viruses including COVID-19

Exclusion Criteria:

- History of prematurity with gestational age at birth less than 35 weeks
- Infants aged ≥22 days with bronchiolitis
- Underlying conditions that affect immunity or may otherwise increase risk of SBI
- Toxic/Septic appearance
- Receiving antibiotic treatment for FWLS
- Routine vaccinations given within the previous 48 hours
- Presenting with seizures
- · Requiring intensive care management
- Identified focus of infection (e.g., cellulitis, acute otitis media in infants >28 days old)

Differential Diagnosis:

Meningitis

Bone and joint infections

Pneumonia

Urinary tract infection

Sepsis/Bacteremia

Enteritis

Herpes Simplex Virus (HSV) infection

Enterovirus

Parechovirus

SARS-CoV-2 Virus

Toxic Criteria (6,7)

Infants that meet ANY of the toxic criteria should receive a full sepsis workup and be admitted to the inpatient area for antibiotic therapy and observation (See Tables 1 & 2).

Signs/Symptoms include:

Poor perfusion

Capillary refill time >2 seconds

Cyanosis

Lethargy

Unable to console

Tachypnea or bradypnea

Hypothermia (96.8°F/36°C)

Table 1. Signs and Symptoms of Shock (8,9)

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	Sign and/or Symptom	
Peripheral Pulses	Decreased or weak Bounding	
Capillary refill	≥ 3 sec Flash (< 1 sec)	
Skin	Mottled, cool Flushed, ruddy, erythroderma (other than face) Petechiae below the nipple, any purpura	
Mental status	Decreased, irritability, confusion inappropriate crying or drowsiness, poor interaction with parents, lethargy, diminished arousability, obtunded	

 $^{^{\}star} \! \uparrow \! HR$ followed by $\! \downarrow \! HR$ with BP changes will be noted as shock becomes uncompensated.

Table 2. Vital Sign Changes of Sepsis (8,9)

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Age	Heart Rate	Respiratory Rate	Systolic BP	
0d - 1m	>205	>60	<60	
>1m to 3m	>205	>60	<70	

†BP changes are late signs of worsening condition. May also present with chills.

<u>Diagnostic Evaluation</u>: In this age group, bacterial pathogens associated with FWLS may include Gram-positive organisms (such as group B Streptococcus, Enterococcus, group A Streptococcus, Staphylococcus aureus, Listeria monocytogenes) and Gram-negative organisms (such as Escherichia coli, Enterobacter, Klebsiella). Streptococcus pneumoniae is more likely to occur in infants >30 days old. (10.11)

Viral pathogens, such as enterovirus, adenovirus, herpes simplex virus, influenza virus, and parainfluenza virus, are also a concern in this patient population. (12,13) There is limited research available on the incidence of SARS-CoV-2 in infants, however preliminary data shows that children, including infants, seem to range from asymptomatic to moderate disease severity.(14)

History: Assess for

Onset of fever

Immunization status (15)

Irritability

Poor feeding

Decreased urine output

Exposure to infectious agents

- Other sick contacts/family members
- Maternal fever at time of delivery
- Maternal Group B streptococcal vaginal colonization
- Maternal HSV infection

Physical Examination:

Rectal temperatures are preferred to axillary or other methods of temperature measurements.

A thorough clinical history and physical examination are essential to determine risk of SBI or identify focus of infection.

Recommended Laboratory Tests

Infants 0 to 21 Days

- Complete blood count (CBC) with differential and platelets
- Blood culture[^] (BC) (obtain prior to antibiotic administration)
- Urinalysis (UA) with micro and culture†^
- Procalcitonin
- Lumbar puncture (LP)* [gram stain, culture, cell count and diff, glucose, protein, viral culture] ‡
- Enterovirus CSF PCR if pleocytosis present
- HSV testing (blood, CSF, swab specimens, cultures of vesicles) if HSV risk factors present or worsening condition.
- Consider laboratory values of inflammation including procalcitonin and C-reactive protein (CRP)

Infants 22 to 28 Days

- Complete blood count (CBC) with differential and platelets
- Urinalysis with micro and culture†^
- Blood culture^ (obtain prior to antibiotic administration)
- Procalcitonin
- If inflammatory markers are abnormal, obtain an LP*. Test for gram stain, culture, cell count and diff, glucose, and protein. If pleocytosis present, obtain enterovirus PCR. ‡
- HSV testing (blood, CSF, swab specimens, cultures of vesicles) if HSV risk factors present or worsening condition.

Infants 29 to 60 Days

- Complete blood count (CBC) with differential and platelets
- Urinalysis with micro and culture†^
- Blood culture^ (obtain prior to antibiotic administration)
- Procalcitonin
- If inflammatory markers are abnormal, consider performing LP*. Test for gram stain, culture, cell count and diff, glucose, and protein. If pleocytosis present, obtain enterovirus PCR. ±
- HSV testing (blood, CSF, swab specimens, cultures of vesicles) if HSV risk factors present or worsening condition.

Optional Laboratory Testing

- Stool for culture and presence of WBCs (if diarrhea present)
- Viral diagnostic testing or rapid tests (if respiratory symptoms)
- Chest X-ray (if respiratory symptoms; WBC >20,000/mm³ or ANC >10,000/mm³)
- † Cath (transurethral catheterization) or SPA (suprapubic aspiration)
- ^ Urine culture and blood culture should be performed prior to antibiotic administration
- *LP should be performed prior to antibiotic administration
- ‡ Tube #1 Glucose, protein
 - Tube #2 Cell count & diff, Gram stain & culture

Laboratory Test Values

Abnormal Inflammatory Markers

- Procalcitonin >0.5 ng/mL
- ANC <1000 mm³ OR >4000 mm³
- * If procalcitonin is not available, may use CRP in combination with elevated temperature as inflammatory markers.

CSF WBC/mm³ Value in Febrile Infants Without Evidence of UTI, IBI, HSV, Enterovirus or Traumatic CSF (18)

- Age 1 28 Days
 - o Abnormal >18 WBC per mm³
- Age 29 60 Days
 - Abnormal >9 WBC per mm³
- * Based on absolute white blood cell count. Values not risk stratified. The AAP guideline does not recommend correcting CSF white count or CSF red blood cell count in children with traumatic lumbar punctures.
- punctures.

 * Children with traumatic lumbar punctures should follow the branch of the algorithm for uninterpretable CSF.

Herpes Simplex Virus

Evidence shows that there is an increase of the risk of mortality for disseminated HSV with each day of delayed treatment. (19) National guidelines support the consideration of HSV infection as a causative agent for neonates with fever. (20) Laboratory evaluation for neonatal HSV infections should be performed for those with clinical findings suggestive of HSV infection or a maternal history of HSV. (21-24)

HSV Risk Factors (23)

- Maternal primary HSV infection
- Maternal fever
- Vaginal delivery
- Prematurity
- Neonatal seizures
- Vesicular rash
- CSF pleocytosis (monocytosis)
- Elevated hepatic enzymes

Signs/Symptoms of Systemic HSV (24)

- Skin, eye, and mouth lesions/disease
- · Seizures, lethargy, and fever
- Disseminated form neonate presents with multi-organ failure

The laboratory tests below are recommended if HSV suspected. (20)

- Specimens of skin vesicles for HSV culture or PCR
- CSF sample for HSV PCR
- Whole blood sample for HSV PCR
- Swab specimens from the mouth, nasopharynx, conjunctivae, and anus for HSV culture (completed inhospital) or PCR (currently a send-out lab)
 - To collect swab specimens for culture, the practitioner may utilize one swab for the eye, mouth, nose and rectum in stated order. Different swabs may be used for each specimen location; however, if this method is used the practitioner should put all swabs in the same transport tube.

Critical Points of Evidence

Evidence Supports

- Complete HSV testing and administer empiric acyclovir for neonates with no identified bacterial pathogen in CSF and the presence
 of CSF pleocytosis and/or exam, concern or possible maternal history of HSV, and/or toxic appearance. (19,21,25-28) Strong
 recommendation, low quality evidence
 - Remarks: Neonates with age less than or equal to 21 days present a heightened concern for HSV.
- Enterovirus testing (if pleocytosis present) should be utilized in addition to usual care in order to decrease length of stay. (29,30) —
 Strong recommendation, low quality evidence
- Enterovirus CSF PCR should be used for testing when CSF specimen is available. (*This recommendation is also based on rapid turnaround time of CSF PCR as compared to serum PCR at TCH.) (31-34) Strong recommendation, low quality evidence
- The diagnostic work-up for neonatal HSV infection should include all of the laboratory tests listed below. (20)
 - Specimens of skin vesicles for HSV culture or PCR
 - CSF sample for HSV PCR
 - Whole blood sample for HSV PCR
 - Swab specimen from the mouth, nasopharynx, conjunctivae, and anus for HSV culture (completed in-house) or PCR (currently a send-out lab)
 - Remarks: To collect swab specimens for culture, the practitioner may utilize one swab for the eye, mouth,
 nose and rectum in stated order. Different swabs may be used for each specimen location; however, if this
 method is used the practitioner should put all swabs in the same transport tube.

Evidence Against

AST and/or ALT lab tests should not routinely be used for screening for disseminated HSV in all infants 0-28 days with fever. (24,25,35-38) – Strong recommendation, low quality evidence

Recommendations Adopted/Adapted from National Guidelines (39)

Well-Appearing Infants 0-21 Days

- · Should assess inflammatory markers.
- Should obtain CSF for analysis (WBC, protein, glucose, Gram stain) and culture for bacteria.
- Should initiate parenteral antimicrobial therapy.
- Clinicians should discontinue parenteral antimicrobial agents and discharge hospitalized patients when all of the following criteria are met: Culture results are negative for 24 to 36 hours or only positive for contaminants; The infant continues to appear clinically well or is improving (e.g., fever, feeding); and There are no other reasons for hospitalization.

Well-Appearing 22-to-28 Day Old Infants

- Should assess inflammatory markers.
- Should obtain CSF for analysis (WBC, protein, glucose, gram stain) and bacterial culture if any inflammatory marker obtained is
 positive.
- Clinicians should administer parenteral antimicrobial therapy in a hospital if either of the following apply: CSF analysis suggests bacterial meningitis; or Urinalysis result is positive.
- Clinicians may administer parenteral antimicrobial therapy in a hospital if all of the following apply: CSF analysis is normal; Urinalysis is normal; and any IM obtained is abnormal.
- If inflammatory markers and UA are normal, should admit without lumbar puncture and observe off parenteral antibiotic therapy. **Remarks:** If there is a clinical concern, may consider LP and admit on antibiotics.
- Clinicians should discontinue antimicrobial agents and discharge hospitalized infants after 24 to 36 hours of negative culture results if both of the following are met: the infant is clinically well or improving (e.g., fever, feeding); there are no other reasons for hospitalization and there is no other infection requiring treatment (e.g., otitis media).

Well-Appearing 29-to-60 Day Old Infants

- Should assess inflammatory markers.
- May obtain CSF for analysis ([WBC, differential, protein, glucose, gram stain], culture for bacteria, and test for enterovirus when CSF pleocytosis is detected) if any inflammatory marker is abnormal.
- Need not to obtain CSF for analysis and culture if all inflammatory markers obtained are normal.
- Clinicians should hospitalize infants if CSF analysis, if obtained, is abnormal.
- Clinicians may hospitalize infants if any inflammatory marker obtained is abnormal.
- Clinicians should manage patients at home if all of the following criteria are met:
 - CSF analysis, if CSF obtained, is normal;
 - urinalysis is negative;
 - all IMs obtained are negative;
 - · appropriate parental education has been provided;
 - follow-up plans for reevaluation in 24 hours have been developed and are in place; and
 - plans have been developed and are in place in case of change in clinical status, including means of communication between family and providers and access to emergency medical care.
- Clinicians may manage infants without antimicrobial treatment at home without having obtained interpretable CSF if all of the following are met: urinalysis is negative; all IMs obtained are normal; and parents can return promptly if there is a change in infant condition and agree to follow-up in 24 hours. Infants monitored at home should be reassessed in the following 24 hours.

Condition-Specific Elements of Clinical Management

Treatment Recommendations (39)

Abnormal Inflammatory Markers

- Procalcitonin >0.5 ng/mL
- ANC <1000 mm³ OR >4000 mm³
- * If procalcitonin is not available, may use CRP in combination with elevated temperature as inflammatory markers.

Neonates (≤21 days): (39)

If presenting in clinic setting, refer to EC.

Evaluate with a full sepsis workup and admit to the inpatient area for antibiotic therapy and observation. (20,21,25-28,31,32-34)

Complete HSV testing and administer empiric acyclovir for neonates with no identified bacterial pathogen in CSF and the presence of CSF pleocytosis or uninterpretable CSF and/or exam, concern or possible maternal history of HSV, and/or toxic appearance. (20,21,25-28) Neonates with age less than or equal to 21 days present a heightened concern for HSV.

Empiric antibiotic therapy of ampicillin and gentamicin should be initiated on all neonates. If there is a concern for meningitis or CSF pleocytosis, ampicillin and cefTAZidime should be administered. (40,41)

Infants 22 to 28 days: (39)

Assess inflammatory markers.

If any inflammatory marker abnormal, obtain an LP.

If urine culture positive or CSF indicates bacterial meningitis, administer antibiotics.

Observe all patients in hospital.

Discontinue antibiotics and discharge hospitalized infants after 24 to 36 hours of negative culture results if both of the following are met:

- Clinically well or improving
- No other reasons for hospitalization and/or no other infection requiring treatment

Infants ≥ 29 days: (39)

Should assess inflammatory markers.

Need not to obtain CSF for analysis and culture if all inflammatory markers obtained are normal.

Obtain CSF for analysis ([WBC, differential, protein, glucose, gram stain], culture for bacteria, and test for enterovirus when CSF pleocytosis is detected) if any inflammatory marker is abnormal.

Admit infants if CSF analysis, if obtained, is abnormal or uninterpretable.

Manage patients at home if all of the following criteria are met:

- CSF analysis, if CSF obtained, is normal;
- urinalysis is negative;
- all IMs obtained are negative;
- appropriate parental education has been provided;
- follow-up plans for reevaluation in 24 hours have been developed and are in place; and
- plans have been developed and are in place in case of change in clinical status, including means of communication between family and providers and access to emergency medical care.

Follow-up Care

Healthcare provider to follow up on blood and urine cultures (if discharged before 48 hours). Urine and CSF cultures are not automatically read.

Healthcare provider to call lab for CSF culture interpretation prior to discharge.

Follow-up appointment with PCP 12-24 hours post-discharge Return to PCP/EC if worsening symptoms

Inpatient/Observation Discharge Criteria

- · Decreasing fever curves
- Well-appearing with no evolution of signs/symptoms
- Tolerating oral intake and maintaining hydration status
- Reliable phone and transportation
- Parent willingness to observe and communicate changes in condition
- Reliable follow-up available 12-24 hours post-discharge
- · Caregiver and PCP agree with plan
- Caregiver understands discharge education

Differences from AAP Guideline

The American Academy of Pediatrics (AAP) Evaluation and Management of Well-Appearing Febrile Infants 8 to 60 Days Old guideline was contextualized to the requirements of patients within Texas Children's Hospital. Due to the concern of distinguishing bronchiolitis from upper respiratory tract infections as well as the risk of a potential untreated SBI, patients with bronchiolitis 21 days or less are included within this guideline. Patients born at 35 weeks gestational age are included within the guideline to provide a standardized treatment path. The gestational age of 35 weeks aligns with care in the newborn nursery as opposed to needing NICU level care. Patients 22-to-28 days old will be observed in the hospital due to the risk of an untreated SBI in our population of patients that includes a large proportion without access to reliable follow-up or a primary care physician. The decision to list procalcitonin and ANC as inflammatory markers was due to availability of procalcitonin at TCH campuses and the better diagnostic accuracy to detect bacterial infection compared to other inflammatory markers. The cut-off value for ANC was adapted locally and a minimum cut-off was added to ensure evaluation for neutropenia. Evidence regarding the diagnostic accuracy of procalcitonin and ANC cut-off values can be found within the AAP Evaluation and Management of Well-Appearing Febrile Infants 8 to 60 Days Old Guideline.

<u>Measures</u>

Outcome

- · Length of stay
- # of readmissions for same problem
- Type of follow-up post EC or Inpatient discharge (phone call vs. visit to PCP)
- # of infants >28 days with LP vs. no LP based on risk criteria
- EC treatment plan for infants after LP performed vs. infants without LP performed
- # of call backs for positive blood cultures
- # of call backs for positive urine cultures for patients with negative UA
- # of enterovirus positive patients with concomitant serious bacteria infection

Table 5. Antibiotic Dose Administration Table (42) Consider insurance/Medicaid formulary restrictions Infants 0 - 21 Days for ≥35 Weeks Gestational Age (GA)

Drug	Dosing Guidelines
Empirical Parenteral Therapy (IV)	
Ampicillin	≤7 days: 100 mg/kg/DOSE every 8 hours
	>7 days:
	GA 35-43 weeks: 50 mg/kg/DOSE every 8 hours
	GA ≥44 weeks: 50 mg/kg/DOSE every 6 hours
	If concern for meningitis: 75 mg/kg/DOSE every 6 hours
Gentamicin	Neonates:
Sulfate	PNA ≤7 days: 4 mg/kg/DOSE every 24 hours
	PNA >7 days: 5 mg/kg/DOSE every 24 hours
Use in lieu of g	entamicin for suspected meningitis or CSF pleocytosis
CefTAZidime	Neonates 0-28 days: 50 mg/kg/DOSE every 8 hours
Treatment of cl	noice for suspected Staphylococcus (10)
Vancomycin	<7 days old:
	>2 kg: 10 to 15 mg/kg/DOSE every 8 to 12 hours
	≥7 to 28 days:
	>2 kg: 10 to 15 mg/kg/DOSE every 6 to 8 hours
Treatment of ch	noice for Herpes Simplex Virus (HSV)
Acyclovir	Neonates and Infants:
i	20 mg/kg/DOSE every 8 hours

Infants 22 - 28 Days

Drug	Dosing Guidelines
Empirical Thera	py (IM/IV)
CefTRIAXone	50 mg/kg/DOSE once
Treatment for Su	uspected Bacterial Meningitis (IV)
Ampicillin [†]	Meningitis or other severe infection, IV: 75 mg/kg/DOSE every 6 hours
CefTRIAXone [†]	Infants 22-28 days: Meningitis, IV: 100 mg/kg/DOSE every 24 hours NOTE: Not for use in patients receiving Y-site administration of calcium-containing IV fluids with a single lumen or single IV site
Treatment of choice for suspected Staphylococcus (10)	
Vancomycin	15 mg/kg/dose every 8 hours

Infants 29 - 60 Davs

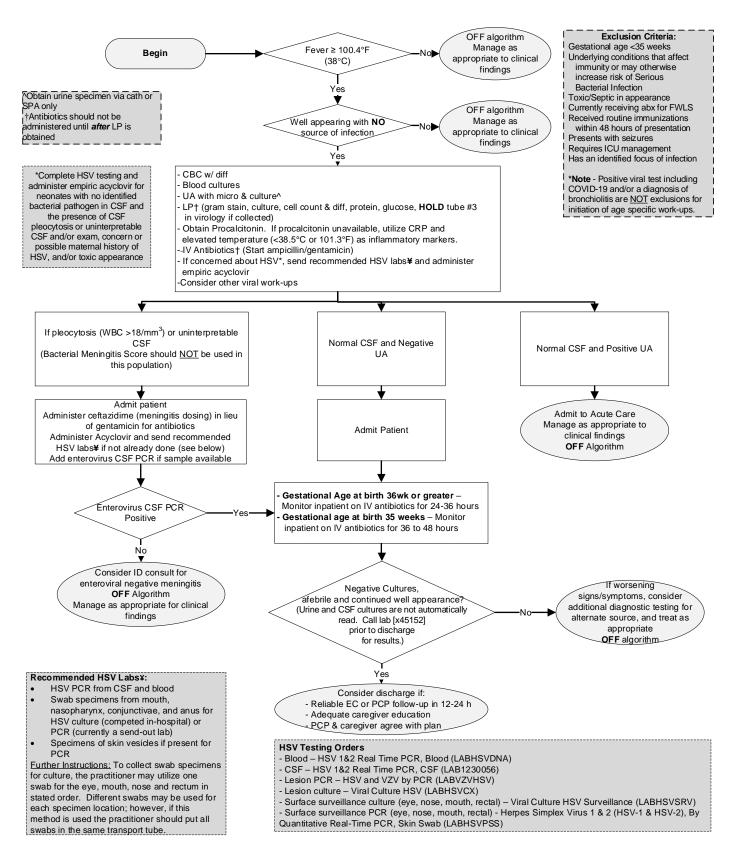
illiants 23 – 00 Days	
Drug	Dosing Guidelines
Outpatient or Er	nergency Center Empirical Therapy (IM/IV)
CefTRIAXone	Infants >28 days: 50 mg/kg/DOSE once
Treatment for S	uspected Bacterial Meningitis (IV)
Vancomycin	Infants >28 days and children:
-	15 mg/kg/dose every 8 hours
CefTRIAXone [†]	Infants >28 days:
	Meningitis, IV: 100 mg/kg/DOSE every 24 hours
	NOTE: Not for use in patients receiving Y-site administration of calcium-containing IV fluids with a single lumen or
	single IV site
Treatment of choice for suspected Staphylococcus (10)	
Vancomycin	15 mg/kg/dose every 8 hours

[†]Reduce antibiotics to general dosing for suspected infection once meningitis ruled out.

^{*}For all age groups, consult ID for patients with enteroviral negative meningitis.

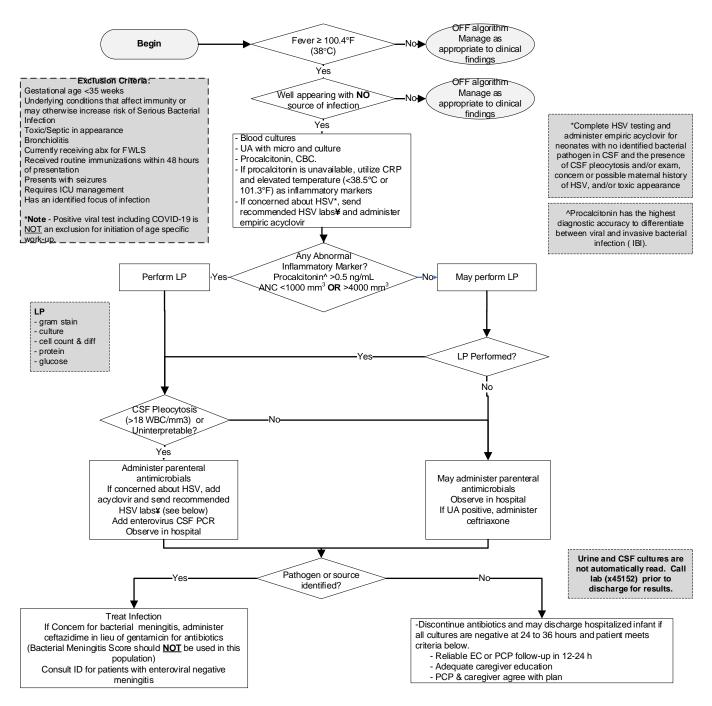
TCH Evidence-Based Outcomes Center Clinical Algorithm Neonates & Infants with Fever Without Localizing Signs (FWLS) 0 - 21 days

Clinical standards are developed for 80% of the patient population with a particular disease. Each practitioner must use his/her clinical judgment in the management of any specific patient.



TCH Evidence-Based Outcomes Center Clinical Algorithm Infants with Fever Without Localizing Signs (FWLS) 22 - 28 days

Clinical standards are developed for 80% of the patient population with a particular disease. Each practitioner must use his/her clinical judgment in the management of any specific patient.



Recommended HSV Labs¥:

- HSV PCR from CSF and blood
- Swab specimens from mouth, nasopharynx, conjunctivae, and anus for HSV culture (competed in-hospital) or PCR (currently a send-out lab)
- Specimens of skin vesicles if present for PCR

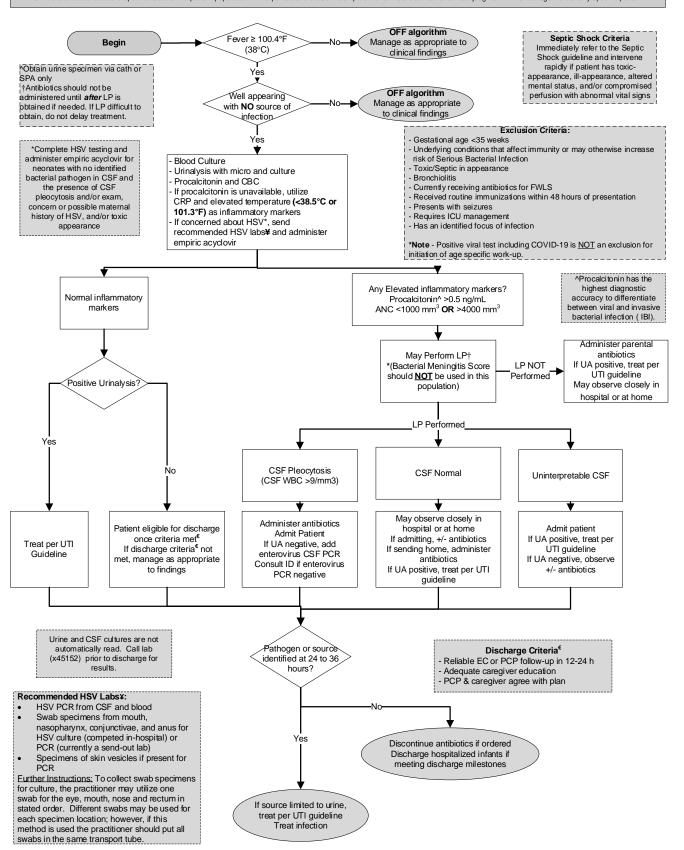
Eurther Instructions: To collect swab specimens for culture, the practitioner may utilize one swab for the eye, mouth, nose and rectum in stated order. Different swabs may be used for each specimen location; however, if this method is used the practitioner should put all swabs in the same transport tube.

HSV Testing Orders

- · Blood HSV 1&2 Real Time PCR, Blood (LABHSVDNA)
- CSF HSV 1&2 Real Time PCR, CSF (LAB1230056)
- Lesion PCR HSV and VZV by PCR (LABVZVHSV)
 Lesion culture Viral Culture HSV (LABHSVCX)
- · Surface surveillance culture (eye, nose, mouth, rectal) Viral Culture HSV Surveillance (LABHSVSRV)
- Surface surveillance PCR (eye, nose, mouth, rectal) Herpes Simplex Virus 1 & 2 (HSV-1 & HSV-2), By Quantitative Real-Time PCR, Skin Swab (LABHSVPSS)

TCH Evidence-Based Outcomes Center Clinical Algorithm Infants with Fever Without Localizing Signs (FWLS) 29 - 60 days

Clinical standards are developed for 80% of the patient population with a particular disease. Each practitioner must use his/her clinical judgment in the management of any specific patient.



References

- 1. Sur, D., & Bukont, E. (2007). Evaluating fever of unidentifiable source in young children. American Family Physician, 75(12), 1805-1811.
- 2. Dagan, R., Sofer, S., Philip, M., & Shachak, E. (1988). Ambulatory care of febrile infants younger than 2 months of age classified as being at low risk for having serious bacterial infections. *Journal of Pediatrics*, 112(3), 355-360.
- 3. Smitherman, H. F., & Macias, C. G. (2019). Febrile infant (younger than 90 days of age): Definition of fever. *UpToDate*. <a href="https://www.uptodate.com/contents/febrile-infant-younger-than-90-days-of-age-definition-of-fever?search=definition-and-etiology-of-fever-in-neonates-and-infants-less-than-&topicRef=6072&source=see_link
- 4. Greenes, D. S. M., & Harper, M. B. M. (1999). Low risk of bacteremia in febrile children with recognizable viral syndromes. *Pediatric Infectious Disease Journal*, 18(3), 258-261.
- 5. Levine, D. A., Platt, S. L., Dayan, P. S., Macias, C. G., Zorc, J. J., Krief, W., et al. (2004). Risk of serious bacterial infection in young febrile infants with respiratory syncytial virus infections. *Pediatrics*, 113(6), 1728-1734.
- 6. Baker, M.D., Bell, L. M., & Avner, J. R. (1993). Outpatient management without antibiotics of fever in selected infants. New England Journal of Medicine, 329(20), 1437-1441.
- 7. Jaskiewicz, J. A., McCarthy, C. A., Richardson, A. C., White, K. C., Fisher, D. J., Powell, K. R., et al. (1994). Febrile infants at low risk for serious bacterial infection--an appraisal of the Rochester criteria and implications for management. *Pediatrics*, *94*(3), 390-396.
- 8. Goldstein, B. M. D., Giroir, B. M. D., Randolph, A. M. D., & Members of the International Consensus Conference on Pediatrics. (2005). International pediatric sepsis consensus conference: Definitions for sepsis and organ dysfunction in pediatrics*. *Pediatric Critical Care Medicine*, 6(1), 2-8.
- 9. American Heart Association. (2012). Pediatric Advanced Life Support (PALS) Provider Manual. Channing Bete Company.
- 10. Fortunov, R. M., Hulten, K. G., Hammerman, W. A., Mason, E. O., Jr., & Kaplan, S. L. (2006). Community-acquired staphylococcus aureus infections in term and near-term previously healthy neonates. *Pediatrics*, *118*(3), 874-881.
- 11. Smitherman, H. F., & Macias, C. G. (2021). Febrile infant (younger than 90 days of age): Outpatient evaluation. *UpToDate*. <a href="https://www.uptodate.com/contents/febrile-infant-younger-than-90-days-of-age-outpatient-evaluation?search=definition-and-etiology-of-fever-in-neonates-and-infants-less-than-&topicRef=106744&source=see_link
- 12. Bilavsky, E., Shouval, D., Yarden-Bilvasky, H., Fisch, N., Ashkenazi, S., & Amir, J. (2008). A prospective study of the risk for serious bacterial infections in hospitalized febrile infants with or without bronchiolitis. *Pediatric Infectious Disease Journal*, 27(3), 269-270.
- Marom, R., Sakran, W., Antonelli, J., Horovitz, Y., Zarfin, Y., Koren, A., et al. (2007). Quick identification of febrile neonates with low risk for serious bacterial infection: An observational study. Archives of Disease in Childhood, Fetal and Neonatal Edition, 92(1), F15-F18.
- 14. DeRose, D., Piersigilli, Ronchetti, M., Santisi, A., Bersani, I., Dotta, A., Danhaive, O., et al. (2020). Novel coronavirus disease (COVID-19) in newborns and infants: What we know so far. *Italian Journal of Pediatrics*, 46(1), 56.
- Baker, M. D., Bell, L. M., & Avner, J. R. (1999). The efficacy of routine outpatient management without antibiotics of fever in selected infants. Pediatrics, 103(3), 627-631.
- 16. Bonsu, B., & Harper, M. (2003). Identifying febrile young infants with bacteremia: Is the peripheral white blood cell count an accurate screen? *Annals of Emergency Medicine*, 42(2), 216-225.
- 17. Garra, G., Cunningham, S. J., & Crain, E. F. (2005). Reappraisal of criteria used to predict serious bacterial illness in febrile infants less than 8 weeks of age. Academy of Emergency Medicine, 12(10), 921-925.
- 18. Byington, C., Kendrick, J., & Sheng, X. (2011). Normative cerebrospinal fluid profiles in febrile infants. Journal of Pediatrics, 158(1), 130-134.
- 19. Shah, S.S., Aronso, P.L., Mohamad, Z. & Lorch, S.A. (2011). Delayed acyclovir therapy and death among neonates with herpes simplex virus infection. *Pediatrics*, 128(6), 1153-1160.
- 20. Committee of Infectious Diseases; American Academy of Pediatrics, Kimberlin, D., Brady, M., Jackson, M., A., & Long, S. (2018). Red book® 2018-2021 report of the committee on infectious diseases (31st ed.). American Academy of Pediatrics.
- 21. Caviness, A. C., Demmler, G. J., Swint, J. M., & Cantor, S. B. (2008). Cost-effectiveness analysis of herpes simplex virus testing and treatment strategies in febrile neonates. *Archives of Pediatrics & Adolescent Medicine*, *162*(7), 665-674.
- 22. Kimberlin, D. W. (2004). Herpes simplex virus, meningitis and encephalitis in neonates. Herpes, 11(Supplement2), 65A-76A.
- 23. Kimberlin, D. W., Lin, C.-Y., Jacobs, R. F., Powell, D. A., Frenkel, L. M., Gruber, W. C., et al. (2001). Natural history of neonatal herpes simplex virus infections in the acyclovir era. *Pediatrics*, 108(2), 223-229.
- 24. Caviness, A. C., Demmler, G. J., & Selwyn, B. J. (2008). Clinical and laboratory features of neonatal herpes simplex virus infection: A case-control study. *Pediatric Infectious Disease Journal*, 27(5), 425-430.
- 25. Long, S.S., Pool, T.E., Vodzak, J., Daskalaki, I. & Gould, J.M. (2011). Herpes Simplex virus infection in young infants during 2 decades of empiric acyclovir therapy. *The Pediatric Infectious Disease Journal*, 30(7), 556-561.
- McGuire, J.L., Zorc, J., Licht, D., Hodinka, R.L., & Shah, S.S. (2012). Herpes simplex testing in neonates in the emergency department. Pediatric Emergency Care, 28(10), 949-955.
- 27. Cruz, A., Freedman, S., Kulik, D., Okada, P., Fleming, A., et al. (2018). Herpes simplex virus infection in infants undergoing meningitis evaluation. *Pediatrics*, 141(2), 1-9.
- 28. VanderPluym, C., Tawfik, G., Hervas-Malo, M., Lacaze-Masmonteil, T., Kellner, J., & Robinson, J. (2012). Empiric acyclovir for neonatal herpes simplex virus infection. *The Journal of Maternal-Fetal & Neonatal Medicine*, *25*(8), 1278-1282.
- 29. Dewan, M., Zorc, J. J., Hodinka, R. L., & Shah, S. S. (2010). Cerebrospinal fluid enterovirus testing in infants 56 days or younger. *Archives of Pediatrics and Adolescent Medicine*, 164(9), 824-30.
- 30. Stellrecht, K. A., Harding, I., Woron, A. M., Lepow, M. L., & Venezia, R. A. (2002). The impact of an enteroviral RT-PCR assay on the diagnosis of aseptic meningitis and patient management. Journal of Clinical Virology, 25, S19-S26.
- 31. Rittichier, K. R., Bryan, P. A., Bassett, K. E., Taggart, E. W., Enriguez, F. R., Hillyard, D. R., et al. (2005). Diagnosis and outcomes of enterovirus infections in young infants. Pediatric Infectious Disease Journal, 24(6), 546-550.
- Byington, C. L., Taggart, W., Carroll, K. C., & Hillyard, D. R. (1999). A polymerase chain reaction- based epidemiologic investigation of the incidence of nonpolio enteroviral infections in febrile and afebrile infants 90 days and younger. Pediatrics, 103(3), e27.
- 33. Ahmed, A., Brito, F., Goto, C., Hickey, S. M., Olsen, K. D., Trujillo, M., et al. (1997). Clinical utility of the polymerase chain reaction for diagnosis of enteroviral meningitis in infancy. *Journal of Pediatrics*, 131(3), 393-397
- 34. Carroll, K. C., Taggart, B., Robison, J., Byington, C., & Hillyard, D. (2000). Evaluation of the Roche AMPLICOR enterovirus PCR assay in the diagnosis of enteroviral central nervous system infections. *Journal of Clinical Virology*, 19(3), 149-156.
- 35. Fidler, K., Pierce, C., Cubitt, W., Novelli, V., & Peters, M. (2004). Could neonatal disseminated herpes simplex virus infections be treated earlier? Journal of Infection, 49, 141-146.
- 36. O'Riordan, D., Golden, W., & Aucott, S. (2006). Herpes simplex virus infection in preterm infants. Pediatrics, 118, e1612-e1620.

- 37. Kotzbaurer, D., Frank, G., Dong, W., & Shore, S. (2014). Clinical and laboratory characteristics of disseminated herpes simplex virus infection in neonates. *Hospital Pediatrics*, *4*, 167-171.
- 38. Wolfert, S., de Jong, E., Vossen, A., Zwaveling, J., Te Pas, A., Walther, F., & Lopriore, E. (2011). Diagnostic and therapeutic management for suspected neonatal herpes simplex virus infection. *Journal of Clinical Virology, 51*, 8-11.
- 39. Pantell, R., Roberts, K., Adams, W., Dreyer, B., Kuppermann, N., O'Leary, S., Okechukwu, K., Woods, C., Subcommittee of Febrile Infants. (2021). Evaluation and management of well-appearing febrile infants 8 to 60 days old. *Pediatrics, 148*(2), e2021052228.
- 40. Byington, C.L., Rittichier, K.E., Bassett, H.C., Glasgow, T.S., & Pavia, A.T. (2003). Serious bacterial infections in febrile infants younger than 90 days of age: the importance of ampicillin-resistant pathogens. *Pediatrics*, 111(5), 964-968.
- 41. Clark, R.H., Bloom, B.T., Spitzer, A.R., & Gerstmann, D.R. (2006). Empiric use of ampicillin and cefotaxime, compared with ampicillin and gentamicin, for neonates at risk for sepsis is associated with an increased risk of neonatal death. *Pediatrics*, 117(1), 67-74
- 42. Texas Children's Hospital Drug Information and Formulary. (2022). Lexicomp Online. http://online.lexi.com/lco/action/home

Clinical Standards Preparation

This clinical standard was prepared by the Evidence-Based Outcomes Center (EBOC) team in collaboration with content experts at Texas Children's Hospital. Development of this clinical standard supports the TCH Quality and Patient Safety Program initiative to promote clinical standards and outcomes that build a culture of quality and safety within the organization.

Fever Without Localizing Signs 0-60 Days Content Expert Team

Joseph Allen, MD, Emergency Medicine Andrea Cruz, MD, Emergency Medicine Charlyn Davis, RN, Emergency Medicine Nursing James Dunn, PhD, Pathology Mindy Fein, MD, Emergency Medicine Catherine Foster, MD, Infectious Disease Lakshmi Katakam, MD, Neonatology Lucilia Marquez, MD, Infectious Disease Brent Mothner, MD, Pediatric Hospital Medicine Aderonke Ojo, MD, Emergency Medicine Debra Palazzi, MD, Infectious Disease Minal Patel, MD, Neonatology Sunjeev Patel, MD, Texas Children's Pediatrics Autumn Pruette, MD, Texas Children's Pediatrics Brittany Rodriguez, PharmD, Pharmacy Cynthia Toy, PharmD, Pharmacy Joe Tran, MD, Pediatric Hospital Medicine Sowdhamini Wallace, MD, Pediatric Hospital Medicine Elizabeth Wuestner, RN, Emergency Center Nursing

EBOC Team

Andrea Jackson, MBA, RN Binita Patel, MD, Chief Medical Quality Officer

Additional EBOC Support

Betsy Lewis, MSN, RN, CNL, Evidence-Based Practice Specialist Sheesha Porter, MSN, RN, Evidence-Based Practice Specialist Anne Dykes, MSN, RN, ACNS-BC, Manager

The following financial and/or intellectual conflict(s) was/were identified and addressed to ensure objectivity: Content Expert Team member A. Cruz, MD is the author of research on clinical decision rules and management of HSV.

Development Process

This clinical standard was developed using the process outlined in the EBOC Manual. The literature appraisal documents the following steps:

- 1. Review Preparation
 - PICO questions established
 - Evidence search confirmed with content experts
- 2. Review of Existing External Guidelines
 - Management of Infants 0-60 Days with Fever of Unknown Source, Cincinnati Children's Hospital; Neonatal Fever, Seattle Children's Hospital; Evaluation/Treatment of Febrile Young Infants (0-56 Days), Children's Hospital of Philadelphia; Red Book – Report of Committee of Infectious Disease, American Academy of Pediatrics
- 3. Literature Review of Relevant Evidence
 - Searched: Medline, Cochrane, Cinahl, AAP, BMJ Clinical Evidence, Google Scholar
- 4. Critically Analyze the Evidence
 - 2 meta-analyses, 41 non-randomized studies, and 4 review articles
- 5. Summarize the Evidence
 - Materials used in the development of the clinical standard, literature appraisal, and any order sets are maintained in a FWLS 0-60 Days electronic manual within EBOC.

Evaluating the Quality of the Evidence

Published clinical guidelines were evaluated for this review using the **AGREE II** criteria. The summary of these guidelines are included in the literature appraisal. AGREE II criteria evaluate Guideline Scope and Purpose, Stakeholder Involvement, Rigor of Development, Clarity and Presentation, Applicability, and Editorial Independence using a 4-point Likert scale. The higher the score, the more comprehensive the guideline.

This clinical standard specifically summarizes the evidence *in support of* or *against* specific interventions and identifies where evidence is *lacking/inconclusive*. The following categories describe how research findings provide support for treatment interventions. *"Evidence Supports"* provides evidence to support an intervention

"Evidence Against" provides evidence against an intervention. "Evidence Lacking/Inconclusive" indicates there is insufficient evidence to support or refute an intervention and no conclusion can be drawn from the evidence.

The **GRADE** criteria were utilized to evaluate the body of evidence used to make practice recommendations. The table below defines how the quality of the evidence is rated and how a strong versus weak recommendation is established. The literature appraisal reflects the critical points of evidence.

Recommendation	
STRONG	Desirable effects clearly outweigh undesirable effects or vice versa
WEAK	Desirable effects closely balanced with undesirable effects
Quality	Type of Evidence
High	Consistent evidence from well-performed RCTs or exceptionally strong evidence from unbiased observational studies
Moderate	Evidence from RCTs with important limitations (e.g., inconsistent results, methodological flaws, indirect evidence, or imprecise results) or unusually strong evidence from unbiased observational studies
Low	Evidence for at least 1 critical outcome from observational studies, RCTs with serious flaws or indirect evidence
Very Low	Evidence for at least 1 critical outcome from unsystematic clinical observations or very indirect evidence

Recommendations

Practice recommendations were directed by the existing evidence and consensus amongst the content experts. Patient and family preferences were included when possible. The Content Expert Team and EBOC team remain aware of the controversies in the management of FWLS in infants 0-60 days. When evidence is lacking, options in care are provided in the clinical standard and the accompanying order sets (if applicable).

Approval Process

Clinical standards are reviewed and approved by hospital committees as deemed appropriate for its intended use. Clinical standards are reviewed as necessary within EBOC at Texas Children's Hospital. Content Expert Teams are involved with every review and update.

Disclaimer

Practice recommendations are based upon the evidence available at the time the clinical standard was developed. Clinical standards (guidelines, summaries, or pathways) do not set out the standard of care and are not intended to be used to dictate a course of care. Each physician/practitioner must use his or her independent judgment in the management of any specific patient and is responsible, in consultation with the patient and/or the patient's family, to make the ultimate judgment regarding care.

Version History

Date	Comments
Mar 2009	Originally completed
Sep 2014	Update
Jul 2015	Revision
Jun 2016	Revision
Feb 2017	Revision
Feb 2021	Update
June 2021	Revision
Feb 2021	Update
April 2022	Update
July 2023	Algorithm (0 – 21 Days) Revised